ORIGINAL PAPER

W. Qian · X. Chen · D. Fu · J. Zou · J. Meng

Intersubgenomic heterosis in seed yield potential observed in a new type of *Brassica napus* introgressed with partial *Brassica rapa* genome

Received: 18 March 2004 / Accepted: 10 January 2005 / Published online: 2 April 2005 © Springer-Verlag 2005

Abstract This paper reports the observation on the intersubgenomic heterosis for seed yield among hybrids between natural Brassica napus (AⁿAⁿCⁿCⁿ) and a new type of B. napus with introgressions of genomic components of Brassica rapa (A^rA^r). This B. napus was selected from the progeny of B. napus \times B. rapa and (B. $napus \times B$. $rapa) \times B$. rapa based on extensive phenotypic and cytological observation. Among the 129 studied partial intersubgenomic hybrids, which were obtained by randomly crossing 13 lines of the new type of B. napus in F₃ or BC₁F₃ to 27 cultivars of B. napus from different regions as tester lines, about 90% of combinations exceeded the yield of their respective tester lines, whereas about 75% and 25% of combinations surpassed two elite Chinese cultivars, respectively. This strong heterosis was further confirmed by reevaluating 2 out of the 129 combinations in a successive year and by surveying hybrids between 20 lines of the new type of B. napus in BC₁F₅ and its parental B. napus in two locations. Some DNA segments from B. rapa were identified with significant effects on seed yield and yield components of the new type of B. napus in BC₁F₅ and intersubgenomic hybrids in positive or negative direction. It seems that the genomic components introgressed from B. rapa contributed to improvement of seed yield of rapeseed.

Communicated by H.C. Becker

W. Qian · X. Chen · D. Fu · J. Zou · J. Meng (☒) National Key Laboratory of Crop Genetic Improvement, Huazhong Agricultural University,

Wuhan 430070, China

E-mail: jmeng@mail.hzau.edu.cn

Tel.: +86-27-87282457 Fax: +86-27-87281006

W. Qian Institute of Crop Science and Plant Breeding, Christian-Albrechts-University of Kiel, 24118 Kiel, Germany **Keywords** Brassica napus · Brassica rapa · Intersubgenomic heterosis · Introgression · Seed yield

Introduction

Brassica napus (AACC, 2n=38) originated from a spontaneous hybridization between B. rapa (AA, 2n=20) and B. oleracea (CC, 2n=18) (UN 1935), containing the entire chromosome sets of both parental genomes (Olsson 1960; Schiemann 1932; Sinskaya 1928; Tsunoda 1980). Although this amphidiploid species was domesticated only about 400 years ago, it became the most important oilseed Brassica crop in the world due to good production potential and resistances (Gómez-Campo 1999; Liu 2000). In China, B. napus accounts for about 85% of oilseed Brassica (Fu 2000). However, modern B. napus cultivars have a narrow genetic basis, limiting its potential for improving seed yield (Becker et al. 1995).

B. rapa, an old oilseed crop, exhibits wide genetic diversity and interesting agronomic traits (Downey and Röbbelen 1989; Prakash and Hinata 1980) and is rather different from B. napus with regard to its genome structure (Hoenecke and Chyi 1991; Song et al. 1988, 1995). To depict those differences, the concept of subgenomes was proposed where "A" was suggested to represent the genome of B. rapa (A'A') and "A" and "C" were defined as the genomes of B. napus (A'A'C'C'C) (Qian et al. 2003).

Intervarietal heterosis has been wildly utilized, and positive correlations between genetic distance between parents of hybrid and mid-parent heterosis has been demonstrated for seed yield in *B. napus* (Ali et al. 1995; Diers et al. 1996; Grant and Beversdorf 1985; Lefort-Buson et al. 1987; Riaz et al. 2001). Due to the large genetic differences between *B. rapa* and *B. napus* and the strong heterosis for biomass yield among hybrids between *B. napus* and *B. rapa* (Liu 2000; Liu et al. 2002; Qian et al. 2003; Sun 1943; Zhao and Becker 1998), a way to utilize intersubgenomic heterosis was proposed

for seed production by crossing natural *B. napus* with a new type of *B. napus* with introgression of the *B. rapa* genome (Liu et al. 2002).

The objectives of this study were (1) to describe the selection of new type of *B. napus* by screening the progeny of interspecific hybrids between *B. napus* and *B. rapa* based on morphological and cytological observations, (2) to exploit the seed yield potential of intersubgenomic hybrids between natural and new type of *B. napus*, and (3) to identify DNA segments introgressed from *B. rapa* with significant effects on seed yield and yield components in the new type of *B. napus* and hybrids derived from them.

Materials and methods

Development of plant material

Fifty of 120 triploid combinations (A^rAⁿCⁿ) between *B. napus* and *B. rapa* with high biomass yield (Liu et al. 2002; Qian et al. 2003) were employed to develop the new type of *B. napus* by two breeding programs: successive selfing them, and backcrossing them to parental *B. rapa* followed by successive selfing (Fig. 1).

The seed yield potential of the new type of B. napus was evaluated by developing hybrids between natural and new type of B. napus in the different generations (Fig. 1), which were defined as partial intersubgenomic hybrids (PIGH) in order to distinguish the A^r subgenome-contained hybrids from the conventional intervarietal hybrids. Individual plants from 13 lines of the new type of B. napus in F₃ and BC₁F₃, as donors of pollen (Table 1), were randomly crossed with 27 cultivars of B. napus from Australia, China, and Europe as tester lines to produce 129 PIGH to test seed yield. In a successive year, the heterosis of PIGH was reevaluated with two PIGH from the new type of B. napus in F_4 , $971 \times XD$ and $77101 \times XD$ (971 and 77101 as test lines and XD as the new type of B. napus derived from a natural B. napus, 'Xiangyou 13'). Moreover, respective intervarietal hybrids (971 × 'Xiangyou 13', 77101 × 'Xiangyou 13') were also produced as additional controls. Twenty inbred lines with a high degree of uniformity but large variation among the lines were chosen from a population of BC₁F₅ derived from an individual of the new type of B. napus in BC₁F₂, H3T1 [('Huashuang 3' × 'Tianmen Youcai') × 'Tianmen Youcai'], to investigate seed yield and yield components as well as the PIGH between them and 'Huashuang 3'. PIGH produced with the new type of B. napus lines at F_3 or BC₁F₃, F₄, and BC₁F₅ generations were named as PIGH-3, PIGH-4, and PIGH-5, respectively.

Field experiments

Randomized complete block design was used for all three field experiments. Plant density was according to farmers' practice in the region of the Yangtze River, i.e., row spacing of 25.6 cm and 12 plants grown in 1.9-m short rows or 25 plants grown in 4-m long rows. As checks, 'Zhongyou 821' (an elite open pollination cultivar with high erucic acid and high glucosinolates in seed) as CK₁ in all field experiments and 'Huaza 4' (an elite commercial canola hybrid) as CK₂ in the field experiment evaluating PIGH-3 and PIGH-4 were chosen, because they have been widely planted in China for several years. Another elite commercial canola hybrid, 'Huaza 6', which has won the first place in the region of Yangtze River in recent years, was used as CK₃ in the field experiment evaluating PIGH-5.

The PIGH-3 were grown with two replications together with tester lines in Wuhan in 2001. CK₁ and CK₂ were planted every ten plots. Due to the limited seeds, plots consisted of single short rows. Ten plants in the

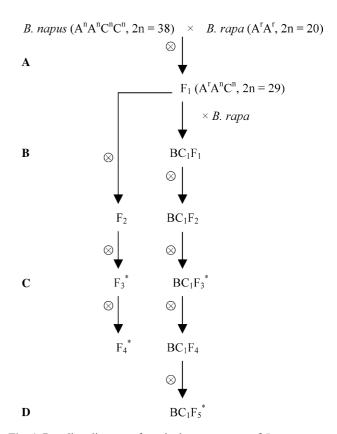


Fig. 1 Breeding diagram of producing a new type of Brassica napus for evaluating intersubgenomic heterosis. A Evaluation of intersubgenomic heterosis among 120 triploid hybrids between B. napus and B. rapa for biomass yield (Liu et al. 2002; Qian et al. 2003). B Selection for the new type of B. napus with 38 chromosomes in somatic cells, high fertility, and A subgenome recombined between Aⁿ and A^r from the progenies of the 50 triploid hybrids with high biomass yield. C Evaluation of heterosis for seed yield among 129 partial intersubgenomic hybrids (PIGH-3) generated with the new type B. napus, and reevaluation of two combinations out of 129 PIGH-3 by developing PIGH-4. D Survey on heterosis among the hybrids between the 20 inbred lines of BC₁F₅and their parental B. napus in two locations and identification of DNA segments from A^rsignificantly affecting seed yield and seed-related traits by amplified fragment length polymorphisms. Asterisks indicate the donors of pollen to produce respective PIGH

Table 1 Characteristics of 29 individual plants of the new type of *Brassica napus* in BC_1F_2 or F_2 and seed yield potential among 129 partial intersubgenomic hybrids (*PIGH*)-3 derived from the ran-

dom crosses between 13 lines of the new type of *B. napus* in F_3 or BC_1F_3 and 27 cultivars of natural *B. napus*. $I(A^r)$ Index of A^r subgenome components

New type	e of B. napus	PIGH-3				
Code	Pedigree	<i>I</i> (A ^r) (%)	Seeds set (seeds/pod)	Number of tester lines	Seed yield (g/plant)	
					Means	Range
BZ2	BC ₁ F ₂ [('Bullet' × 'Zhejiang 1') × 'Zhejiang 1']	60.2	12.8			
H3M1	BC ₁ F ₂ [('Huashuang 3' × 'Maverick') × 'Maverick']	68.2	12.6	17	13.4 ^a	8.2-21.2
H3M2	BC_1F_2 [('Huashuang 3' × 'Maverick') × 'Maverick']	43.6	11.2			
H3M3	BC_1F_2 [('Huashuang 3' × 'Maverick') × 'Maverick']	61.8	9.6			
H3M4	BC_1F_2 [('Huashuang 3' × 'Maverick') × 'Maverick']	55.4	11.6			
H3T1	BC ₁ F ₂ [('Huashuang 3' × 'Tianmen Youcai') × 'Tianmen Youcai']	69.6	13.0	9	10.2	7.3–12.7
Н3Т2	BC ₁ F ₂ [('Huashuang 3' × 'Tianmen Youcai') × 'Tianmen Youcai']	63.4	9.0			
H2T1	BC ₁ F ₂ [('Huashuang 2' × 'Tianmen Daye Youcai') × 'Tianmen Daye Youcai']	67.7	14.0	9	10.9	4.9–14.0
H2T2	BC ₁ F ₂ [('Huashuang 2' × 'Tianmen Daye Youcai') × 'Tianmen Daye Youcai']	54.2	9.8			
Mean	•	60.5	11.5			
6D1	F_2 (6203 × 'Denglong Zhong')	56.0	15.8	13	10.6	4.8 - 16.3
6D2	F_2 (6203 × 'Denglong Zhong')	52.0	13.4	9	11.1	6.3 - 13.8
6Y1	F_2 (6203 × 'Yanyou 2')	61.5	18.0	11	14.7 ^{a,b}	12.7 - 17.5
6Y2	F_2 (6203 × 'Yanyou 2')	45.2	9.0			
6Z	F_2 (6203 × 'Zhejiang1')	28.2	7.0			
BD	F_2 ('Bullet' × 'Denglong Zhong')	56.7	8.0	11	10.4	3.7 - 15.0
BX1	F ₂ ('Bullet' × 'Xishui Youcai Bai')	54.6	9.0	9	10.0	6.7 - 12.8
BX2	F ₂ ('Bullet' × 'Xishui Youcai Bai')	65.7	7.6			
BX3	F ₂ ('Bullet' × 'Xishui Youcai Bai')	33.3	6.6	8	7.9	5.2 - 11.8
BX4	F ₂ ('Bullet' × 'Xishui Youcai Bai')	54.5	11.2			
BZ1	F ₂ ('Bullet' × 'Zhejiang 1')	56.7	13.0			
BT	F ₂ ('Bullet' × 'Tianmen Daye Youcai')	_	9.0			
H2X	F ₂ ('Huashuang 2' × 'Xishui Youcai Bai')	56.1	7.2	8	11.4	5.7 - 15.8
H2X	F ₂ ('Huashuang 2' × 'Xishui Youcai Bai')	_	7.0			
H3Y	F ₂ ('Huashuang 3' × 'Yanyou 2')	53.2	14.0			
SZ	F_2 (S2501 × 'Zhejiang1')	48.6	16.0	5	12.9 ^a	10.6-15.2
ST	F ₂ (S2501× 'Tianmen Youcai')	50.0	12.6			
SY	F ₂ (S2501× 'Yaanhuang')	35.2	14.0			
XC	F ₂ ('Xianyou 13' × 'Chengdu Ai Youcai')	52.3	9.0	11	9.2	2.8-12.4
XD	F ₂ ('Xianyou 13' × 'Denglong Zhong')	35.7	10.4	6	11.9	10.1 - 14.9
Mean		49.3	10.9			

^aSignificant exceeding check cultivar 1 (CK₁), 'Zhongyou 821', at P = 0.05

center of each plot were harvested to calculate the seed yield per plant.

Seeds of PIGH-4 were sown together with respective intervarietal hybrids and tester lines with three replications in Wuhan in 2002. One hundred and twenty-five plants were grown per plot with five long rows, and all plants in the plot were harvested to measure seed yield.

Seeds of PIGH-5 and their parents were sown with three replications in two locations, Wuhan and Jingzhou, in 2002. Every plot was composed of 36 individual plants in three short rows. Ten plants in the center of each plot were harvested to calculate the seed yield per plant and yield components.

Cytological examination

The number of chromosomes in somatic cells of candidate individuals of the new type of *B. napus* was counted

according to the method of Li et al. (1995). Ten to thirty metaphase cells from young buds were evaluated in each plant. Chromosomes were stained with 10% of carbol fuchsin.

Molecular marker assay

Amplified fragment length polymorphism (AFLP) markers were developed from DNA samples of the new type of B. napus and their parents, following the method of Vos et al. (1995) and Xu et al. (2001). Five and 19 pairs of primers were used to detect the DNA introgression from B. rapa among the new type of B. napus in F_2 – BC_1F_2 and BC_1F_5 populations, respectively. The presence or absence of an AFLP band was scored with 1 or 0, respectively. The degree of introgression from B. rapa was described with the index of A^r subgenome

bSignificant exceeding check cultivar 2 (CK₂), 'Huaza 4', at P = 0.05

components in the new type of B. napus, symbolized as $I(A^r)$, which was calculated as follows:

$$I(A^r) = n_A^r/N \times 100$$

where n_A^r represents the number of the AFLP bands that were present in both the new type of *B. napus* and parental *B. rapa*, but absent in parental *B. napus*, and *N* represents the total number of polymorphic AFLP bands in its parental *B. napus* and *B. rapa*. The genetic distance (GD) between line X and Y was calculated on the basis of their Dice genetic similar coefficient (Nei and Li 1979) as follows:

$$GD_X y = 1 - 2N_X y/(N_X + N_Y)$$

where N_{XY} is the number of common bands shared by lines X and Y, and N_X and N_Y are the total number of bands in line X and Y, respectively.

The effects of polymorphic AFLP molecular marker loci on seed yield and yield components were assessed by one-way analysis of variance (ANOVA) in the inbred lines of BC₁F₅ and PIGH-5, respectively (Zhang et al. 1994; Liu et al. 2002). If a molecular marker locus had significant effects on a trait, we denoted it as an active marker locus. If the allele from A^r in an active marker locus had significant positively or negative effect, the locus was defined as a favorable active marker locus (FAM) or an unfavorable active marker locus (UAM), respectively. The effect value of an active marker locus was estimated as the phenotype difference between the average of inbred lines or hybrids from inbred lines that had the same band as the B. rapa cultivar 'Tianmen Youcai' and the average of the other inbred lines or hybrids from inbred lines that had the same band as 'Huashuang 3' at an active marker locus.

The ANOVA was done with the Statistical Analysis System (SAS Institute 1992). Pearson's simple correlation coefficients were calculated among variables of interest.

Results

Introgression of the genetic components of B. rapa

The triploid F_1 plants ($A^rA^nC^n$, 2n=29) exhibited low fertility, with an average of 4.4 seeds/pod among the 120 interspecific combinations between *B. napus* and *B. rapa* (data not shown). On the other hand, some individuals from F_2 and BC_1F_2 restored the fertility to some extent. There were 29 individual plants of the new type of *B. napus* with 38 chromosomes and high fecundity selected from the population of F_2 and BC_1F_2 (Table 1). Out of these, 20 plants were identified from 1,872 individuals in F_2 , and nine plants were identified from 860 individuals in BC_1F_2 , which were the descendants of four individuals with 29 chromosomes selected from 482 BC_1F_1 plants. It seems that recombined euploid gametes (AC,

n=19) were highly abundant in the generations of F_1 , BC_1F_1 , and BC_1F_2 .

Introgression from A^r in the new type of B. napus was identified with AFLP markers. The I(A^r) of 29 individuals in F₂ and BC₁F₂ varied from 28.2% to 69.6%, with the average of 60.5% for BC₁F₂ and 49.3% for F₂ estimated with about 100 polymorphic markers (Table 1). When identified 20 inbred lines of BC₁F₅ using 344 polymorphic markers, the $I(A^{r})$ of this population was 42.3% on average, ranging from 38.8% to 45.0%, which was obviously lower than that of its ancestor, H3T1, with the $I(A^r)$ of 69.6% (Tables 1, 2). Moreover, recombination might happen in the process of the introgression, because we detected about 6% of new bands differed from their parents' band in the 20 inbred lines of BC₁F₅. However, the genetic distance between the inbred line of BC₁F₅ and 'Huashuang 3' significantly and positively related with the $I(A^{r})$ $(r=0.67, P \le 0.01)$. In other words, the larger the introgression from A^r in the new type of B. napus, the greater the genetic distance between it and its parental B. napus.

Partial intersubgenomic heterosis potential for seed yield

Strong seed yield heterosis was observed among PIGH in all three field experiments. Among 129 PIGH-3, about 90% of combinations exceeded their respective tester lines whereas about 75% and 25% of combinations surpassed CK_1 and CK_2 , respectively (Fig.2). It should be noted that some of new type of *B. napus* lines, for example, 6Y1, H3M1, and SZ, exhibited high combining ability for seed yield, because the average seed yield of combinations derived from them significantly exceeded that of the controls ($P \le 0.05$) (Table 1).

Two combinations, $971 \times \text{XD}$ and $77101 \times \text{XD}$, which slightly exceeded CK₁ and CK₂, respectively, in the field trial evaluating PIGH-3, still exhibited obvious heterosis in PIGH-4 in the enlarged area of plot in a successive year. They exceeded the respective tester line, controls, and intervarietal hybrid in seed yield, and the combination, $77101 \times \text{XD}$, even significantly exceeded the controls ($P \le 0.05$) (Fig.3).

The mean value of accessions over two locations in the field trial evaluating PIGH-5 is shown in Table 2. Seed yield showed the highest level of heterosis, followed by pods per plant and seeds per pod; very little heterosis was detected for seed weight. All PIGH-5 exceeded 'Huashuang 3', with heterosis from 29.17% to 95.83%, and the amount of mid-parental heterosis varied from 21.73% to 86.50%, with an average of 43.15% for seed yield. Some inbred lines and PIGH-5 significantly exceeded controls on seed yield and yield components ($P \le 0.05$). Joint ANOVA between two locations showed significant differences for seed yield and yield components ($P \le 0.01$) and no significant interactions between accessions and locations (Table 3).

Table 2 Performances of seed yield and yield components (means \pm SD) among PIGH-5 derived from the inbred lines of BC₁F₅, their parents, and the controls over two locations

Code	I(A ^r)	Pods/plant		Seeds/pod		Weight of 1,000 seeds (g)		Seed yield (g/plant)	
		Inbred lines	Hybrids	Inbred lines	Hybrids	Inbred lines	Hybrids	Inbred lines	Hybrids
1 2 3 4 5 6 7 8 9 10 11 12 13	42.6 43.4 41.9 43.0 42.6 43.8 43.0 42.2 41.9 38.8 44.2 44.6 40.3	221.3 ± 32.3 174.5 ± 22.1 184.1 ± 47.1 192.9 ± 20.4 260.2 ± 40.3 204.6 ± 22.7 231.4 ± 16.3 207.6 ± 35.1 182.8 ± 34.9 229.4 ± 80.5 213.2 ± 46.1 240.6 ± 51.3 156.1 ± 28.7	276.9 ± 131.8^{a} 218.8 ± 32.3 247.4 ± 51.5 204.2 ± 33.5 263.0 ± 39.5^{a} 272.2 ± 48.9^{a} 246.5 ± 53.9 $292.3 \pm 56.8^{a,b}$ 209.6 ± 15.0 $282.3 \pm 31.3^{a,b}$ $282.3 \pm 37.6^{a,b}$ 212.0 ± 26.5 188.5 ± 35.2	16.0 ± 2.8 16.0 ± 2.5 16.4 ± 3.1 18.4 ± 4.1 14.3 ± 3.3 15.2 ± 2.9 19.2 ± 1.5 17.5 ± 3.0 17.8 ± 0.9 16.0 ± 1.2 17.0 ± 4.2 11.3 ± 4.0 19.8 ± 4.0	17.2 ± 3.7 18.1 ± 4.0 18.0 ± 2.7 19.2 ± 2.9 16.8 ± 1.2 18.3 ± 2.8 17.4 ± 1.5 17.5 ± 1.3 19.0 ± 2.5 16.5 ± 2.8 17.2 ± 2.7 19.1 ± 3.2 19.0 ± 1.4	$\begin{array}{c} 3.8 \pm 0.2^{a,b} \\ 3.7 \pm 0.3^{a,b} \\ 3.8 \pm 0.3^{a,b} \\ 3.8 \pm 0.2 \\ 3.7 \pm 0.4^{a,b} \\ 3.7 \pm 0.2^{a,b} \\ 3.7 \pm 0.2^{a,b} \\ 3.9 \pm 0.3^{a,b} \\ 3.9 \pm 0.2^{a,b} \\ 3.5 \pm 0.3^{b} \\ 3.6 \pm 0.4^{a,b} \\ 4.1 \pm 0.3^{a,b} \\ 3.4 \pm 0.1 \end{array}$	$\begin{array}{c} 3.4\pm0.4\\ 3.6\pm0.2^{a,b}\\ 4.0\pm0.2^{a,b}\\ 3.6\pm0.2^{a,b}\\ 3.5\pm0.2^{b}\\ 3.8\pm0.2^{a,b}\\ 3.9\pm0.2^{a,b}\\ 3.9\pm0.2^{a,b}\\ 3.7\pm0.2^{a,b}\\ 3.7\pm0.2^{a,b}\\ 3.3\pm0.1\\ 3.9\pm0.3^{a,b}\\ 3.8\pm0.3^{a,b}\\ 3.8\pm0.3^{a,b}\\ \end{array}$	8.3 ± 2.1 6.6 ± 0.8 8.1 ± 2.6 9.0 ± 1.8 8.5 ± 1.8 7.7 ± 2.5 $10.6 \pm 0.6^{a,b}$ 8.5 ± 2.0 6.9 ± 1.6 7.3 ± 1.9 7.5 ± 1.2 5.3 ± 1.5 8.1 ± 2.3	$\begin{array}{c} 10.2 \pm 4.1 \\ 9.8 \pm 1.0 \\ 10.3 \pm 2.9 \\ 10.1 \pm 1.7 \\ 9.7 \pm 3.9 \\ 14.1 \pm 2.3^{b} \\ 10.9 \pm 2.6^{a,b} \\ 13.6 \pm 2.4^{b} \\ 10.3 \pm 2.1 \\ 11.6 \pm 1.5^{a,b} \\ 12.3 \pm 2.6^{b} \\ 9.7 \pm 1.3 \\ 10.0 \pm 2.6 \end{array}$
14 15 16 17 18 19 20 Means Huashuang 3 CK ₁ CK ₃	42.2 39.5 38.8 39.5 45.0 45.0 43.4 42.3	$\begin{array}{c} 130.1 \pm 26.7 \\ 208.0 \pm 28.0 \\ 222.5 \pm 57.8 \\ 185.3 \pm 32.2 \\ 203.3 \pm 24.3 \\ 161.7 \pm 32.1 \\ 162.6 \pm 62.1 \\ 196.3 \pm 43.7 \\ 201.9 \\ 217.1 \pm 47.1 \\ 205.2 \pm 33.6 \end{array}$	$\begin{array}{c} 188.3 \pm 33.2 \\ 225.3 \pm 12.1 \\ 263.8 \pm 51.5^{a} \\ 222.7 \pm 32.1 \\ 245.1 \pm 42.2 \\ 194.8 \pm 34.5 \\ 238.5 \pm 48.9 \\ 247.2 \pm 30.9 \\ 241.7 \\ \\ 237.2 \pm 28.6 \end{array}$	19.8 ± 4.0 15.3 ± 4.2 15.1 ± 2.7 16.9 ± 1.9 11.5 ± 1.8 19.2 ± 4.2 15.4 ± 2.4 14.7 ± 2.6 16.1 16.7 ± 1.0 19.4 ± 3.6	19.0 ± 1.4 18.2 ± 1.5 16.0 ± 0.8 19.7 ± 4.0 15.3 ± 1.4 21.1 ± 3.9 15.1 ± 2.3 17.4 ± 2.8 17.8	3.9 ± 0.1 $3.9 \pm 0.3^{a,b}$ $4.1 \pm 0.2^{a,b}$ $3.8 \pm 0.2^{a,b}$ $3.8 \pm 0.1^{a,b}$ 3.3 ± 0.2 $4.0 \pm 0.1^{a,b}$ 3.5 ± 0.1^{b} 3.7 3.4 ± 0.1 3.3 ± 0.1	3.0 ± 0.3 $3.7 \pm 0.2^{a,b}$ $3.7 \pm 0.2^{a,b}$ $3.8 \pm 0.2^{a,b}$ $3.9 \pm 0.2^{a,b}$ $3.8 \pm 0.1^{a,b}$ $4.0 \pm 0.3^{a,b}$ 3.5 ± 0.1^{b} 3.7	8.1 ± 2.3 8.4 ± 1.1 7.8 ± 1.7 7.0 ± 0.9 6.5 ± 1.5 8.0 ± 1.3 6.7 ± 2.0 6.9 ± 2.6 7.7 7.2 ± 0.9 8.3 ± 1.7	10.0 \pm 2.0 a,b 11.1 \pm 1.0 a,b 9.9 \pm 1.8 9.7 \pm 1.5 9.6 \pm 1.9 10.0 \pm 1.7 9.3 \pm 2.3 11.1 \pm 2.2 a,b 10.7

^aSignificant, exceeding Ck_1 at P = 0.05 ^bSignificant, exceeding Ck_2 at P = 0.05

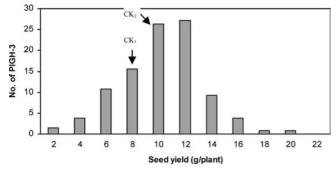


Fig. 2 Frequency distributions of the average of seed yield (g/plant) of 129 PIGH-3. The *arrows* indicate the positions of the two check cultivars, 'Zhongyou 821' (CK_1), and 'Huaza 4' (CK_2).

Identification of DNA segments from A^r with significant effects on seed yield and yield components

Among 344 polymorphic AFLP marker loci detected in the 20 inbred lines of BC_1F_5 , there were 41 active marker loci that had significant effects on seed yield or yield components in the inbred lines and hybrids ($P \le 0.05$), 24 FAMs, and 27 UAMs (Table 4). In addition, six marker loci contributed to two or more of the traits. The total of effects of FAMs exceeded those of UAMs for seed yield and yield components except for pods per plant in the inbred lines and hybrids and seeds per pod in inbred lines.

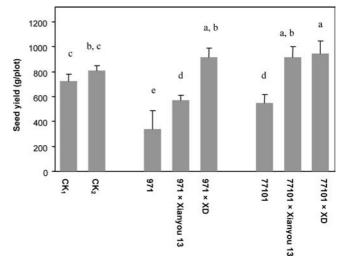


Fig. 3 Comparison of two PIGH-4 with respective tester lines and intervarietal hybrids for seed yield shown as mean values (*columns*) and half the standard deviation (*bars*). Columns with the same letter do not different significantly

Discussion

Introgression of A^r subgenome

Generally, there are two strategies to introgress genetic components of *B. rapa* into *B. napus*. One strategy is to

Table 3 Joint one-way analysis of variance for yield and yield components of the PIGH-5, their parents, and the controls over two locations

Items	df	Mean squares		Seed weight (g/1,000)	Seed yield (g)
		Pods/ plant	Seeds/ pod		
Replications (locations) Locations Accesions	4	13,857.3*	1.48	0.012	17.29*
	1	48,459.2*	447.22*	0.268*	167.53*
	42	7,076.2*	22.26*	0.330*	21.66*
Accesion ^a locations	42	1,351.2	7.12	0.062	2.31
Error	168	1,565.3	5.61	0.052	3.29

Table 4 Active molecular marker loci with significant effects on seed yield and yield components and the effects of the alleles from A^r at those loci on these traits in the 20 inbred lines of BC_1F_5 and PIGH-5

Item	Inbred lines	Hybrids	Overlap	
Pods/plant				
UAM^a	$2(-73.5)^{b}$	8 (-255.7)	1	
FAM ^c	2 (59.7)	3 (124.8)	0	
Sum	4 (-13.8)	11(-130.9)	1	
Seeds/pod	, ,	, ,		
UAM	4 (-12.1)	2(-3.4)	1	
FAM	0 `	3 (5.2)	0	
Sum	4 (-12.1)	5 (1.8)	1	
Seed weight (g/1,000	seeds)	` '		
UAM	4 (-1.0)	1(-0.5)	0	
FAM	4 (1.1)	3 (5.6)	0	
Sum	8 (0.1)	4 (5.1)	0	
Seed yield (g/plant)	• •	. ,		
UAM	2(-5.6)	4(-6.3)	0	
FAM	6 (11.8)	3 (6.4)	0	
Sum	8 (6.2)	7 (0.1)	0	

^aUAM Unfavorable active marker locus ^bNumber of active marker loci, the sum of effects of the alleles from the introgressed A^r ^cFAM Favorable active marker locus

artificially resynthesized B. napus (ArArCoCo) by sexual or somatic hybridization between B. rapa and B. oleracea (C°C°). There were some successes transferring gene(s) (Akbar 1989; Chen et al. 1988; Heath and Earle 1997; Mackay 1977; Morgan et al. 1998; Olsson and Ellerström 1980; Prakash and Raut 1983; Ren et al. 2000: Schranz and Osborn 2000). However, multivalent or univalent formation at meiosis result in low fertility and low seed set in these resynthesized lines, especially in low generations, which were less direct utilized in seed production (Olsson 1960; Schenck and Röbbelen 1982; Prakash and Raut 1983; Sundberg et al. 1987; Rosén et al. 1988; Heath and Earle 1996, 1997). Namai et al. (1980) suggested a way to reduce chromosomal irregularities and to enhance the seeding fertility for the artificially resynthesized B. napus by crossing it with natural B. napus. In the practical breeding program, resynthesized B. napus was usually backcrossed to natural B. napus for several times as bridging materials or raw materials, and only a few favorable genetic components from B. rapa were transferred into B. napus (Kräling 1987; Becker et al. 1995; Prakash et al. 1999). The other strategy is to develop the new type of B. napus from the progeny of hybrid between B. napus and B. rapa. Easy development to hybrids between B. napus and B. rapa and high frequencies of euploid gametes in the F₁ have enabled this strategy to be widely utilized in Asia (Liu 2000; Lu and Masahiro 2001; Mikkelsen et al. 1996; Shiga 1970; Zhou and Scarth 1995). Namai (1976) observed higher frequency of multivalents in the meiosis of triploid with the genomic constitute of A^r A^r C^o than that of A^rAⁿCⁿ. In this study, the observation of high fertility among the new type of *B. napus* in low generations and of high seed yield potential of PIGH also suggests that the A^r/Aⁿ genome pair is more compatible or harmonious with the Cⁿ genome.

Although genomic components of Ar were transferred into the new type of B. napus, it seemed to exist a decreasing tendency to I(A^r) from low to high generations. An important reason was that the candidate individuals in high generations possessed some basic traits partially or completely controlled by the gene(s) in Aⁿ, not in A^r. For example, low erucic acid and low glucosinolates in seed, which were present in parental B. napus 'Huashuang 3' as a canola cultivar, but not in parental B. rapa 'Tianmen Youcai' as an old cultivar. In other words, the direction of artificial selection for these traits in high generations was negative against the introgression of A^r. Moreover, a relatively high degree of heterozygosity in low generations might lead to overestimation of the $I(A^r)$ of low generations, because AFLPs as dominant markers were used.

Intersubgenomic heterosis in B. napus

Intergenomic heterosis is a universal phenomenon in nature, which is extensively and directly utilized in forest

^aSignificant at P = 0.01

tree and grass (Allard 1960; Brewbaker and Sun 1999). Recent advances in heterosis have generated considerable interest in intersubspecific heterosis for seed yield in rice (Yuan 1994, 1997, 1998; Peng et al. 1999). In this study, we observed that some PIGH derived from the new type of *B. napus* in different generations exhibited strong vigor, and detected that the DNA segments introgressed from A^r had positive effects on seed yield on the whole. It suggests that the utilization of intersubgenomic heterosis be an accessible breeding strategy to increase seed yield in rapeseed.

It should be noted that genomic components of A^r were partially transferred into the new type of *B. napus*, and that some DNA segments introgressed from A^r unfavorably took effects on seed yield and yield components in this study. The potential of intersubgenomic heterosis may be enhanced by pyramiding more favorable genomic components of A^r.

Acknowledgements This work was supported by National 863 High Technology Program in China and Special Research Fund for the Doctoral Program of Higher Education. The authors gratefully acknowledge Professor J. Wu for assistance in plant selection. We also thank Dr. Christian Jung, Dr. Martin Frauen, Dr. Jianwei Zhao, and Dr. Jinguo Hu for critically reading the manuscript.

References

- Akbar M (1989) Chromosomal stability and performance of resynthesized *Brassica napus* produced for gain in earliness and short-day response. Hereditas 111:247–253
- Ali M, Copeland LD, Elias SG, Kelley JD (1995) Relationship between genetic distance and heterosis for yield and morphological traits in winter canola (*Brassica napus* L.). Theor Appl Genet 91:118–121
- Allard RW (1960) Principles of plant breeding. Wiley, New York, pp 434-443
- Becker HC, Engqvist GM, Karlsson B (1995) Comparison of rapeseed cultivars and resynthesized lines based on allozyme and RFLP markers. Theor Appl Genet 91:62–67
- Brewbaker JL, Sun WG (1999) Trees and heterosis. In: Coors JG, Pandey S (eds) The genetics and exploitation of heterosis in crops. American Society of Agronomy, Crop Science Society of America, Soil Science Society of America, Madison, pp 463–478
- Chen BY, Heneen WK, Jonsson R (1988) Resynthesis of *Brassica napus* L. through interspecific hybridization between *B. alboglabra* Bailey and *B. campestris* L. with special emphasis on seed colour. Plant Breed 101:52–59
- Diers BW, McVetty PBW, Osborn TC (1996) Relationship between heterosis and genetic distance based on restriction fragment length polymorphism markers in oilseed rape (*Brassica napus* L.). Crop Sci 36:79–83
- Downey RK, Röbbelen G (1989) *Brassica* species. In: Röbbelen G, Downey RK, Ashri A (eds) Oil crops of the world. McGraw-Hill, New York, pp 339–362
- Fu T (2000) Breeding and utilization of rapeseed hybrid. Hubei Science Technology, Hubei, pp 167–169
- Gómez-Campo C, (1999) Biology of *Brassica* coenospecies. Elsevier, The Netherlands, p 49
- Grant I, Beversdorf WD (1985) Heterosis and combining ability estimates in spring-planted oilseed rape (*Brassica napus* L.). Can J Genet Cytol 27:472–478
- Heath DW, Earle ED (1996) Resynthesis of rapeseed (*Brassica napus* L.): a comparison of sexual versus somatic hybridization. Plant Breed 115:395–401

- Heath DW, Earle ED (1997) Resynthesis of low linolenic acid rapeseed (*Brassica napus* L.) through protoplast fusion. Euphytica 93:339–344
- Hoenecke M, Chyi YS (1991) Comparison of *Brassica napus* and *B. rapa* genomes based on restriction fragment length polymorphism mapping. In: Proceedings of the 8th international rapeseed congress, vol 4, Saskatchewan, pp 1102–1107
- Kräling K (1987) Utilization of genetic variability of resynthesized rapeseed. Plant Breed 99:209–217
- Lefort-Buson M, Guillot-Lemoine B, Dattee Y (1987) Heterosis and genetic distance in rapeseed (*Brassica napus* L.): crosses between European and Asiatic selfed lines. Genome 29:413–418
- Li Z, Liu H, Luo P (1995) Production and cytogenetics of intergeneric hybrids between *Brassica napus* and *Orychophragmus violaceus*. Theor Appl Genet 91:131–136
- Liu H (2000) Genetics and breeding in rapeseed. Chinese Agricultural Universitatis, Beijing, pp 144–177
- Liu R, Qian W, Meng J (2002) Association of RFLP markers and biomass heterosis in trigenomic hybrids of oilseed rape (*Brassica napus* × *B. campestris*). Theor Appl Genet 105:1050–1057
- Lu C, Masahiro K (2001) Fertilization fitness and relation to chromosome number in interspecific progeny between *Brassica napus* and *B. comparative* study using current and resynthesized *B. napus*. Breed Sci 51:73–81
- Mackay GR (1977) The introgression of S alleles into forage rape *Brassica napus* L. from turnip, *Brassica campestris* L. ssp. *rapifera*. Euphytica 26:511–519
- Mikkelsen TR, Jensen J, Jørgensen RB (1996) Inheritance of oilseed rape (*Brassica napus*) RAPD markers in a backcross progeny with *Brassica campestris*. Theor Appl Genet 92:492–497
- Morgan CL, Bruce DM, Child R, Ladbrooke ZL, Arthur AE (1998) Genetic variation for pod shatter resistance among lines of oilseed rape developed from synthetic *B. napus*. Field Crops Res 58:153–165
- Namai H (1976) Cytogenetic and breeding studies on the transfer of economic characters by means of interspecific and intergenomic crossing in the tribe Brassiceae of Cruciferae. Mem Fac Agr Tokyo Univ Educ 22:101–171
- Namai H, Sarashima M, Hosoda T (1980) Interspecific and intergeneric hybridization breeding in Japan. In: Tsunoda S, Hinata K, Gómez-Campo C (eds) *Brassica* crop and wild allies. Biology and breeding. Japan Scientific Societies, Tokyo, pp 191–203
- Nei M, Li WH (1979) Mathematical model for studying genetic variation in terms of restriction endonucleases. Proc Natl Acad Sci USA 76:5269–5273
- Olsson G (1960) Species crosses within the genus *Brassica napus* L. II. Artificial *Brassica napus* L. Hereditas 46:351–396
- Olsson G, Ellerström S (1980) Polyploidy breeding in Europe. In: Tsunoda S, Hinata K, Gómez-Campo C (eds) *Brassica* crops and wild allies—biology and breeding. Japan Scientific Societies, Tokyo, pp 167–190
- Peng S, Cassman KG, Virmani SS, Sheehy J, Khush GS (1999) Yield potential trends of tropical rice since release of IR8 and the challenge of increasing rice yield potential. Crop Sci 39:1552–1559
- Prakash S, Hinata K (1980) Taxonomy, cytogenetics and origin of crop Brassicas, a review. Opera Bot 55:3–57
- Prakash S, Raut RN (1983) Artificial synthesis of *Brassica napus* and its prospects as an oilseed crop in India. Indian J Genet 43:283–191
- Prakash S, Takahata Y, Kirti PB, Chopra VL (1999) Cytogenetics. In: Gómez-Campo C (ed) Biology of *Brassica* coenospecies. Elsevier, The Netherlands, pp 59–90
- Qian W, Liu R, Meng J (2003) Genetic effects on biomass yield in interspecific hybrids between *Brassica napus* and *B. rapa*. Euphytica 134:9–15
- Ren JP, Dickson MH, Earle ED (2000) Improved resistance to bacterial soft rot by protoplast fusion between *Brassica rapa* and *B. oleracea*. Theor Appl Genet 100:810–819

- Riaz A, Li Q, Quresh Z, Swati MS, Quiros CF (2001) Genetic diversity of oilseed *Brassica napus* inbred lines on sequencerelated amplified polymorphism and its relation to hybrid performance. Plant Breed 120:411–415
- Rosén B, Halldén C, Heneen WK (1988) Diploid Brassica napus somatic hybrids: characterization of nuclear and organellar DNA. Theor Appl Genet 76:197–203
- SAS Institute (1992) SAS technical report. SAS statistics software: changes and enhancements. Release 6.07. SAS Institute, Cary
- Schenck HR, Röbbelen G (1982) Somatic hybrids by fusion of protoplasts from *Brassica oleracea* and *B. campestris*. Z Pflanzenz 89:278–288
- Schiemann E (1932) Entstehung der Kulturpflanzen. Handb Vererbwis 3:271–288
- Schranz ME, Osborn TC (2000) Novel flowering time variation in the resynthesized polyploid *Brassica napus*. J Hered 91:242–246
- Shiga T (1970) Rapa breeding by interspecific crossing between *Brassica napus* and *Brassica campestris* in Japan. Jpn Agric Res Quart 5:5–10
- Sinskaya EN (1928) The oleiferous plants and root crops of the family Cruciferae. Bull Appl Bot Genet Plant Breed 9:641–648
- Song KM, Osborn TC, Williams PH (1988) *Brassica* taxonomy based on nuclear restriction fragment length polymorphism (RFLP) 1.Genome evolution of diploid and amphidiploid species. Theor Appl Genet 75:784–794
- Song KM, Lu P, Tang K, Osborn TC (1995) Rapid genome change in synthetic polyploids of *Brassica* and its implication for polyploid evolution. Proc Natl Acad Sci USA 92:7719–7723
- Sun VG (1943) Heterosis between *Brassica* species. Zhong Hua Nong Xue Hui Bao 175:35–38
- Sundberg E, Landgren M, Glimelius K (1987) Fertility and chromosome stability in *Brassica napus* resynthesized by protoplast fusion. Theor Appl Genet 75:96–104

- Tsunoda S (1980) Ecophysiology of wild and cultivated froms in *Brassica* and allied genera. In: Tsunoda S, Hinata K, Gómez-Campo C (eds) *Brassica* crop and wild allies. Biology and breeding. Japan Scientific Societies, Tokyo, pp 109–120
- UN (1935) Genomic analysis in Brassica with special reference to the experimental formation of *B. napus* and peculiar mode of fertilization. Jpn J Bot 7:389–452
- Vos P, Hogegers R, Bleeker M, Reijians M, Lee T, Hornes M, Frijters A, Pot J, Peleman J, Kuiper M, Zabeau M (1995) AFLP: a new technique for DNA fingerprinting. Nucleic Acids Res 23:4407–4417
- Xu FS, Wang YH, Meng J (2001) Mapping boron efficiency gene(s) in *Brassica napus* using RFLP and AFLP markers. Plant Breed 120:319–324
- Yuan LP (1994) Increasing yield potential in rice by exploitation of heterosis. In: Virmani SS (ed) Hybrid rice technology, new developments and future prospects. IRRI, Los Banos, Philippines, pp 1–6
- Yuan LP (1997) Hybrid rice breeding for super high yield. Hybrid Rice 12(6):1–6
- Yuan LP (1998) Hybrid rice breeding in China. In: Virmani SS, Siddiq EA, Muralidharan K (eds) Advances in hybrid rice technology. Proceedings of the 3rd international symposium on Hybrid Rice, Hyderabad, India, 14–16 November 1996. IRRI, Los Banos, Philippines, pp27–33
- Zhang Q, Gao YJ, Yang SH, Ragab RA, Saghai MA, Li ZB (1994) A half-diallel analysis of heterosis in elite hybrid rice based on RFLPs and microsatellites. Theor Appl Genet 89:185–192
- Zhao J, Becker HC (1998) Genetic variation in Chinese and European oilseed rape (*B. napus*) and turnip rape (*B. campestris*) analysis with isozymes. Acta Agron Sinica 24:213–220
- Zhou Y, Scarth R (1995) Microspore culture of hybrids between *Brassica napus* and *B. campestris*. Acta Bot Sinica 37:848–855